

**Fig. 6** Wood anatomy of the *Tabebuia* alliance (including *Crescentieae*). **a** *Handroanthus barbatus*, TS, diffuse porous wood, paratracheal aliform parenchyma, predominantly unilateral around certain vessels (*asterisks*), with short confluges, thick-walled fibers, growth rings delimited by a line of marginal parenchyma (*arrows*). **b** *Tabebuia roseoalba*, TS, diffuse-porous wood, aliform parenchyma, sometimes unilateral, with short confluges, growth rings delimited by a band of marginal parenchyma. **c** *Zeyheria montana*, TS, diffuse porous wood, aliform parenchyma with short to long confluges, thick walled fibers, growth rings delimited by a line of marginal

parenchyma (*arrows*). **d** *Crescentia cujete*, TS, diffuse porous wood, abundant aliform parenchyma with long confluges forming bands, growth rings delimited by a line of marginal parenchyma. **e** *Crescentia alata*, LT, axial and radial elements storied, uniseriate rays, sometimes biseriate, two cells per parenchyma strand. **f** *Handroanthus chrysotrichus*, LT, all axial and radial elements storied, biseriate rays, parenchyma paratracheal with two cells per strand, large intervessel pits. **g** *Tabebuia fluviatilis*, LR, homocellular rays. **h** *Crescentia cujete*, LR, abundant axial parenchyma, homocellular rays. *Scale bars* **a, b, d, g, h** 100  $\mu$ m, **c** 500  $\mu$ m, **e, f** 200  $\mu$ m

mm<sup>2</sup>),  $43 \pm 4/\text{mm}^2$  in *Cybastax antisiphilitica*. Deposits of lapachol are common in *Handroanthus* and *Zeyheria*. Perforation plates are simple, sometimes foraminata in *Tabebuia*, *Spirotecoma*, *Crescentia* and *Parmentiera*. Intervessel pits vary from small ( $\sim 5 \mu\text{m}$  in *Roseodendron donnel-smithii*) to large ( $\sim 18 \mu\text{m}$  in *Handroanthus chrysotrichus*). Vessel-ray pits are similar in size and shape to the intervessel pits. Fibers are non-septate, thin- to thick-walled and frequently very thick walled (e.g., *Handroanthus*, *Paratecoma peroba*, *Zeyheria*; Fig. 6a, c). Parenchyma ranges from vasicentric to aliform, forming short (Fig. 6a, b) to long confluges (Fig. 6c, d), usually forming bands (Fig. 6d). Unilateral aliform parenchyma

(Fig. 6a, b) is common in some species (e.g., *Handroanthus albus*, *H. impetiginosus*, *Tabebuia roseoalba*). A gradual change from aliform, to aliform-confluent with short to long confluges and finally a marginal band within each growth ring is found in some members of this clade (*Crescentia alata*, *Handroanthus* and *Tabebuia*). All sampled species of *Amphitecna*, *Cybastax antisiphilitica*, *Crescentia alata* and some species of *Handroanthus* (e.g., *Handroanthus barbatus*, *H. chrysotrichus*, *H. impetiginosus*; Fig. 6e, f) have 2 cells per parenchyma strand, while others have 2–4 cells (e.g., *H. serratifolium*). Rays are usually storied (Fig. 6e, f) or irregularly storied, uniseriate (Fig. 6e) to 2–3-seriate (Fig. 6f), generally homocellular

(Fig. 6g, h), although homocellular and heterocellular rays with one marginal row of square cells are present in *Godmania aesculifolia*, *Sparattosperma leucanthum*, and in both species of *Zeyheria* sampled.

#### Paleotropical clade (including Coleeae)

We sampled 22 of the 139 species currently recognized in the Paleotropical clade, representing 15 out of the 19 genera currently included in this group (Lohmann and Ulloa 2006 onwards); only *Phylloctenium* (ditypic genus from Madagascar), *Dinklageodoxa* (monotypic genus from Liberia), and *Rhigozum* (seven species from Tropical Africa and Madagascar) were not sampled. More specifically, we sampled *Catophractes alexandri* (monotypic), *Colea* (2 of 21 spp), *Dolichandrone* (2 of 10 species), *Fernandoa* (2 of 14 species), *Heterophragma* (both species), *Kigelia africana* (monotypic), *Markhamia* (2 of 10 species), *Newbouldia laevis* (monotypic), *Ophiocolea floribunda* (1 of 5 species), *Pajanelia longifolia* (monotypic), *Phyllarthron bojeranum* (1 of 15 species), *Radermachera* (5 of 18 species), *Rhodocolea* (3 of 7 species), *Spathodea campanulata* (monotypic), *Stereospermum chelonoides* (1 of 20 species), and *Tecomella undulata* (monotypic). Of these, *Colea*, *Ophiocolea*, *Phyllarthron*, and *Rhodocolea* belong to tribe Coleeae, a monophyletic group nested within the Paleotropical clade.

#### Diagnostic features

The wood anatomy of most species from the Paleotropical clade is distinctive in having abundant paratracheal aliform confluent parenchyma, with short to long confluences, medium vessels, non-storied, 2–3-seriate, co-occurring homo and heterocellular rays, and thick walled, non-septate fibers. However, Coleeae differ from the rest of the clade by its apotracheal diffuse axial parenchyma that co-occurs with paratracheal vascentric to aliform parenchyma, thin- to very thick-walled fibers, narrow vessels and homocellular uniseriate rays.

#### Detailed description

*Growth rings* are distinct, delimited by a line of marginal parenchyma associated with very narrow vessels (Fig. 7a, b), vessels of larger diameter (Fig. 7b), thicker-walled and radially flattened fibers (Fig. 7b), and sometimes dilated rays (e.g., *Fernandoa adenophylla*, *Markhamia lutea*, *Tecomella undulata*). *Porosity* is diffuse (Fig. 7a, e, f) or semi-ring porous (Fig. 7b), especially in species growing in more arid areas (e.g. *Catophractes alexandri*). *Vessels* are solitary or in multiples of 2–3 (Fig. 7a, b, e, f),  $1.51 \pm 0.44$

vessels/group, medium ( $110 \pm 20 \mu\text{m}$ ), except in Coleeae, where they are narrow ( $60 \pm 10 \mu\text{m}$ ; Fig. 7f); frequencies are variable, ranging from 8 to  $10/\text{mm}^2$  in *Kigelia africana* and *Spathodea campanulata* to 24–40/ $\text{mm}^2$  in *Tecomella undulata*, *Radermachera glandulosa*, and species of tribe Coleeae. *Intervessel pits* are small (4–5  $\mu\text{m}$ ) to minute in tribe Coleeae (<3  $\mu\text{m}$ ). *Vessel-ray pits* are similar in size and shape to intervessel pits. *Fibers* are thin- (Fig. 7b) to very thick-walled (Fig. 7a, e), with simple pits and occasionally septate. Within tribe Coleeae fibers are thin-walled in *Colea* and *Ophiocolea* and very thick-walled in *Rhodocolea* and *Phyllarthron*. Fibers are always septate in the sampled species of *Heterophragma*, *Radermachera*, and *Pajanelia longifolia*. *Parenchyma* is paratracheal vascentric to aliform, with short (Fig. 7e) to long confluences (Fig. 7a, b). Apotracheal diffuse parenchyma is present in all genera from tribe Coleeae (Fig. 7e, f), less abundantly in *Phyllarthron bojeranum* and *Rhodocolea*, always co-occurring with paratracheal vascentric to aliform parenchyma, with short to long confluences in *Rhodocolea multiflora* and *R. nycteriphilla*. Usually 3–4 cells per parenchyma strand (2 cells in *Spathodea campanulata*). *Rays* are non-storied, 3–4-seriate (Fig. 7c), homocellular (Fig. 7d) and heterocellular with one row of marginal square cells common (Fig. 7c). In representatives of tribe Coleeae rays are exclusively unicellular and homocellular (Fig. 7g, h), similarly to *Dolichandrone atrovirens*, *Kigelia africana*, *Stereospermum leucanthum*, and *Tecomella undulata* (which fall outside Coleeae).

#### Oroxyleae

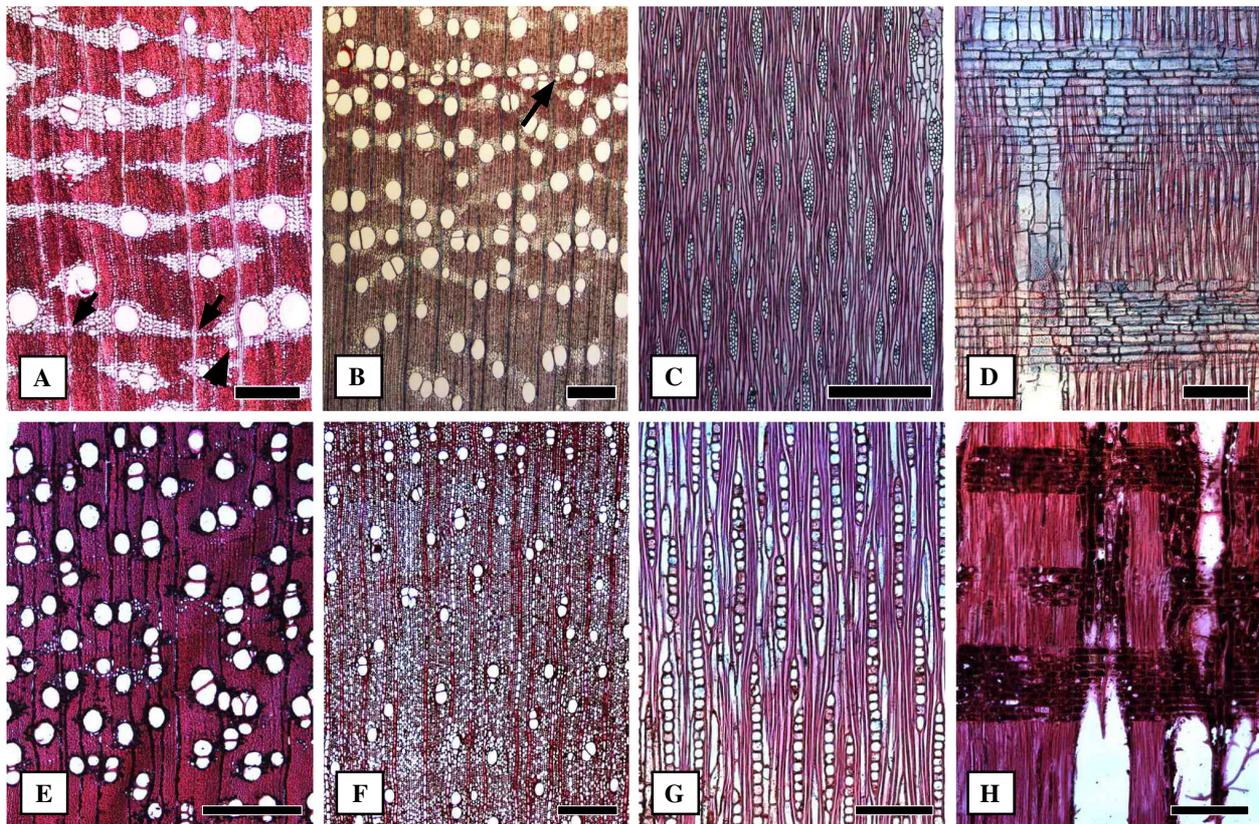
We sampled *Millingtonia hortensis* (monotypic) and *Oroxylum indicum* (monotypic), two of the four genera and six species currently recognized in the tribe (Lohmann and Ulloa 2006 onwards). Only *Nyctocalos* (three species) and *Hieris curtisii* (monotypic), both lianas from Malesia, were not sampled.

#### Diagnostic features

The wood of Oroxyleae can be recognized by a unique combination of vessels with foraminat perforation plates, co-occurring with vessels of simple perforation plates, paratracheal vascentric to aliform parenchyma, homocellular and non-storied rays, fibers generally non-septate.

#### Detailed description

*Growth rings* are distinct, delimited by a line of marginal parenchyma (Fig. 8a), thicker-walled and radially flattened



**Fig. 7** Wood anatomy of the Paleotropical clade (including Coleae). **a** *Stereospermum chelonoides*, TS, diffuse porous wood, vessels solitary to multiples of 2–3, growth rings delimited by a line of marginal parenchyma (arrows) associated with narrow vessels (arrowhead), paratracheal aliform parenchyma with short confluent. **b** *Markhamia lutea*, TS, semi-ring porous wood, vessels solitary to multiples 2–3, growth rings delimited by wide vessels and a line of marginal parenchyma associated with narrow vessels (arrow), aliform parenchyma with short to long confluent. **c** *Spathodea campanulata*, LT, rays 2–3-seriate, non-storied.

**d** *Heterophragma roxburghii*, LR, homocellular rays. **e–h** Tribe Coleae. **e** *Rhodocolea telfairae*, TS, diffuse porous, vessels solitary to multiples of 2–3, paratracheal aliform parenchyma with short confluent, and scanty diffuse apotracheal parenchyma, fibers very thick-walled. **f** *Ophiocolea floribunda*, TS, vessels solitary to multiples of 2–3, abundant apotracheal diffuse parenchyma combined with vascentric paratracheal parenchyma, fibers thin-walled. **g** *Ophiocolea floribunda*, LT, uniseriate rays, non-storied. **h** *Rhodocolea telfairae*, LR, homocellular rays. Scale bars **a**, **c**, **e**, **f** 500  $\mu$ m, **b** 1 mm, **d**, **g**, **h** 200  $\mu$ m

fibers, and dilated rays. Porosity diffuse (Fig. 8a, e). Vessels are solitary or in multiples of 2–3 (Fig. 8a, e),  $1.59 \pm 0.49$  vessels/groups, narrow in *Millingtonia hortensis* ( $80 \pm 26 \mu$ m; Fig. 8a) and medium in *Oroxylum indicum* ( $179 \pm 16 \mu$ m; Fig. 8e);  $25 \pm 8/\text{mm}^2$  of frequency in *Millingtonia hortensis* (Fig. 8a) and fewer,  $4 \pm 4/\text{mm}^2$ , in *Oroxylum indicum* (Fig. 8e). Vessels with foraminiferous perforation plates are common (Fig. 8d, h) co-occurring with vessels with simple perforation plates (more frequent). In *Millingtonia hortensis*, a combination of foraminiferous and reticulate perforation plates is frequently found at the same plate (Fig. 8d). Intervessel pits are small (4–5  $\mu$ m). Vessel-ray pits are similar to the intervessel pits in size and shape. Fibers are thin- to thick-walled (Fig. 8a, e), with simple pits, non-septate to occasionally septate in some individuals of *Oroxylum indicum*. Parenchyma is vascentric (Fig. 8a) to aliform (Fig. 8e), both with short confluent; 3–4 cells per

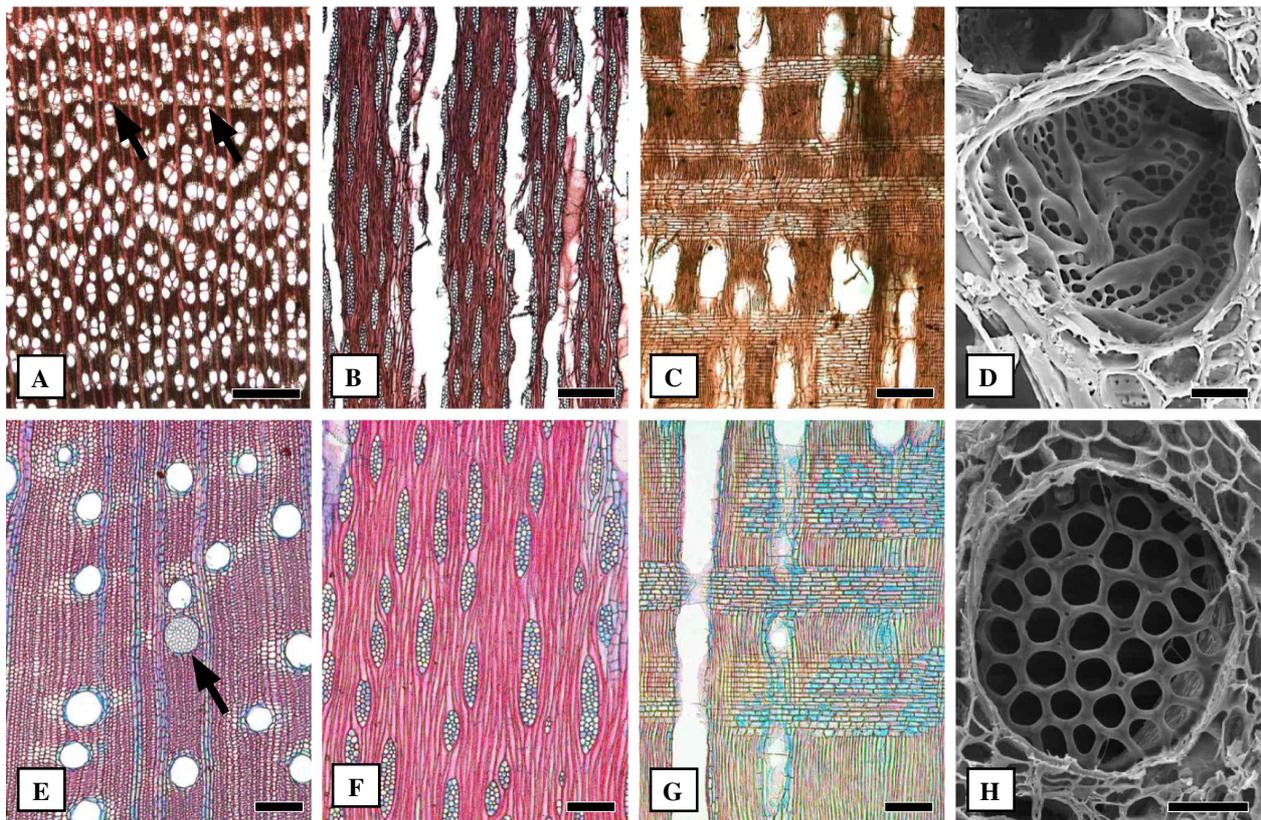
parenchyma strand. Rays are homocellular (Fig. 8c, g), 3-seriate and non-storied (Fig. 8b, f).

### Catalpeae

We sampled *Chilopsis linearis* (monotypic), and *Catalpa* (3 out of the 11), representing the two genera currently included in this clade (Lohmann and Ulloa 2006 onwards). In *Catalpa*, we sampled species from section *Macrocatappa* (*Catalpa longissima*), and section *Catalpa* (*Catalpa bignonioides* and *Catalpa speciosa*).

### Diagnostic features

Catalpeae are distinctive for the semi-ring porous to ring-porous wood found in *Chilopsis linearis* and *Catalpa*



**Fig. 8** Wood anatomy of Oroxyloae. **a–d** *Millingtonia hortensis*. **a** TS, vessels solitary to multiples of two to four, growth ring delimited by a line of marginal parenchyma (arrows) and thicker-walled and radially flattened fibers, vascentric parenchyma, with short confluentes. **b** LT, rays 3-seriate, non-storied. **c** LR, rays homocellular. **d** Scanning electron microscopy (SEM) of a vessel with a combination of foraminiate and reticulate perforation plate together,

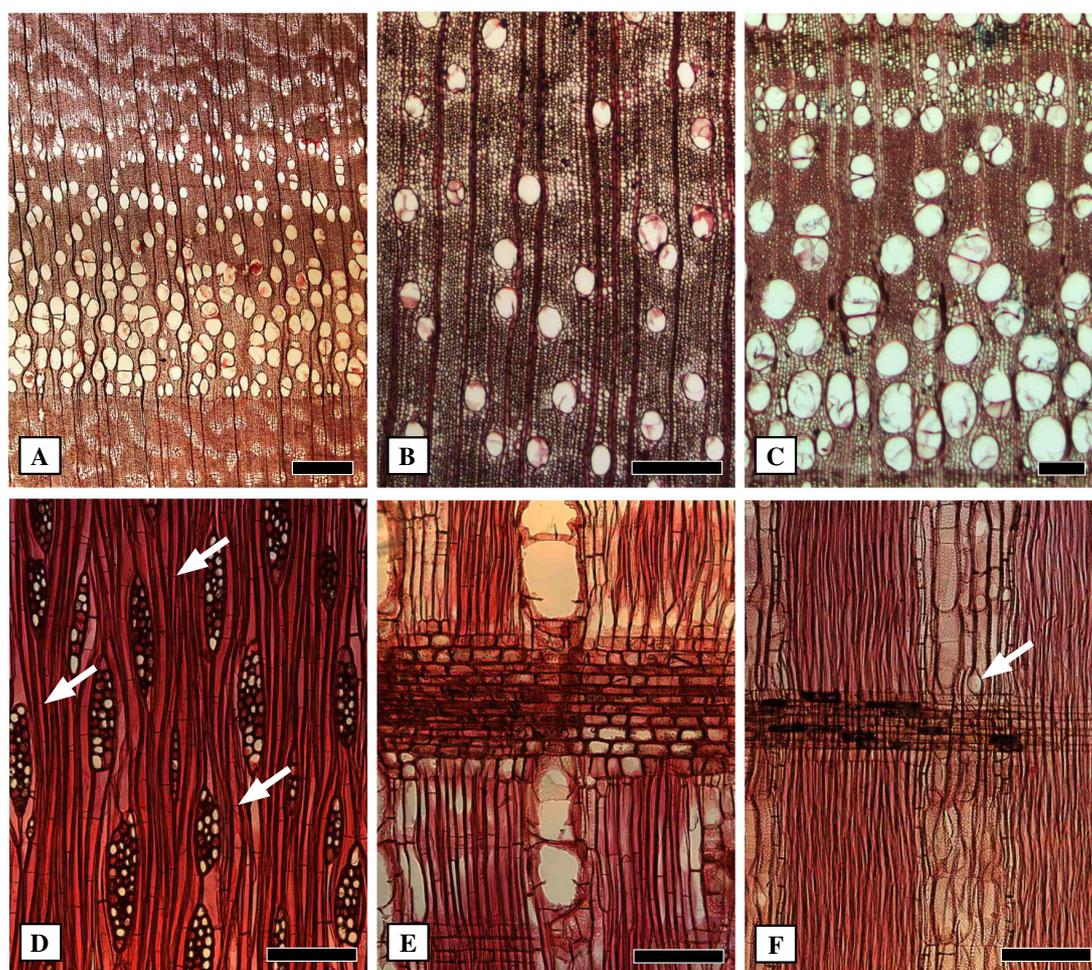
typical of this species. **e, f** *Oroxylum indicum*. **e** TS, solitary vessels common, paratracheal aliform parenchyma with short confluentes, foraminiate perforation plate (arrow). **f** LT, rays 3-seriate, non-storied. **g** LR, rays homocellular. **h** SEM of a vessel with foraminiate perforation plate. Scale bars **a** 500  $\mu\text{m}$ , **b, c, e–g** 200  $\mu\text{m}$ , **d** 20  $\mu\text{m}$ , **h** 50  $\mu\text{m}$

(section *Catalpa*), as well as the presence of scanty paratracheal to vascentric confluent parenchyma. In *Catalpa* (section *Macrocatappa*) the parenchyma is aliform confluent. Tyloses are abundant in both sections, as are the septate fibers in the tropical and subtropical species (*Chilopsis linearis* and *C. longissima*) and the unique presence of simple to semi bordered vessel-ray pits.

*Detailed description*

*Growth rings* are distinct, delimited by a band of marginal parenchyma, vessels of two different diameters in *Chilopsis linearis* (Fig. 9c), *Catalpa speciosa* and *C. bignonioides* (section *Catalpa*; Fig. 9a), thicker-walled and radially flattened fibers. *Porosity* diffuse in *Catalpa longissima* (section *Macrocatappa*; Fig. 9b), and semi-ring porous to ring-porous in *Catalpa speciosa*, *C. bignonioides* (section *Catalpa*; Fig. 9a), and *Chilopsis linearis* (Fig. 9c). *Vessels* are solitary or in multiples of 2–3,  $1.43 \pm 0.12$

vessels/group in earlywood and tropical specimens,  $19.27 \pm 7.59$  vessels/group in latewood, wide vessels with  $200 \pm 18 \mu\text{m}$  in the semi-ring to ring-porous woods (i.e., *Catalpa* section *Catalpa* + *Chilopsis*), and medium vessels with  $131 \pm 10 \mu\text{m}$  in *Catalpa longissima* (section *Macrocatappa*); narrow vessels of latewood of  $30 \pm 6 \mu\text{m}$ ; and frequency of  $12 \pm 5/\text{mm}^2$  in *Catalpa speciosa* and  $34 \pm 10/\text{mm}^2$  in *Chilopsis linearis*. Helical thickening is present in all vessels of *Catalpa speciosa* (section *Catalpa*) and *Chilopsis linearis*. *Perforation plates* are simple (Fig. 9f). *Intervessel pits* are small (5–7  $\mu\text{m}$ ). *Vessel-ray pitting* is simple to semi-bordered. *Fibers* are septate in the tropical and subtropical species (Fig. 9d, e) and non septate in the temperate species (Table 1), thin- to thick-walled, with simple pits. *Parenchyma* is scanty paratracheal to vascentric in *Catalpa speciosa* and *Catalpa bignonioides* (section *Catalpa*) and *Chilopsis*, vascentric forming bands when associated with the narrow vessels in the latewood of *Chilopsis linearis*. Bands of marginal parenchyma are



**Fig. 9** Wood anatomy of Catalpeae. **a** *Catalpa speciosa* (section *Catalpa*), TS, semi-ring porous wood, earlywood vessels solitary to multiples of 2–3, associated with a band of marginal parenchyma, latewood vessels in clusters, scanty paratracheal parenchyma, tyloses common. **b** *Catalpa longissima* (section *Macrocatapa*), TS, wood diffuse porous, vessels solitary to multiples of 2–3, parenchyma aliform, with short confluences, tyloses common. **c** *Chilopsis linearis*, TS, semi-ring porous wood, earlywood vessels solitary to multiples of

2–3, associated with a band of marginal parenchyma, latewood vessels generally in multiples, with paratracheal vasicentric parenchyma forming bands, tyloses common. **d** *Catalpa longissima*, LR, 2–3-seriate rays, non-storied, septate fibers common (arrows). **e** *Chilopsis linearis*, LR, heterocellular rays with one row of square marginal cells. **f** *Catalpa speciosa*, LR, heterocellular ray, narrow vessels in clusters, showing simple perforation plates. Scale bars **a** 500  $\mu\text{m}$ , **b–f** 200  $\mu\text{m}$

present. In *Catalpa longissima* (section *Macrocatapa*), the parenchyma is aliform with short confluences (Fig. 9b). Parenchyma strands have 3–4 cells, as seen in tangential section. Rays are 3-seriate, non-storied (Fig. 9d), and heterocellular with one row of marginal square cells (Fig. 9e, f) co-occurring with homocellular rays.

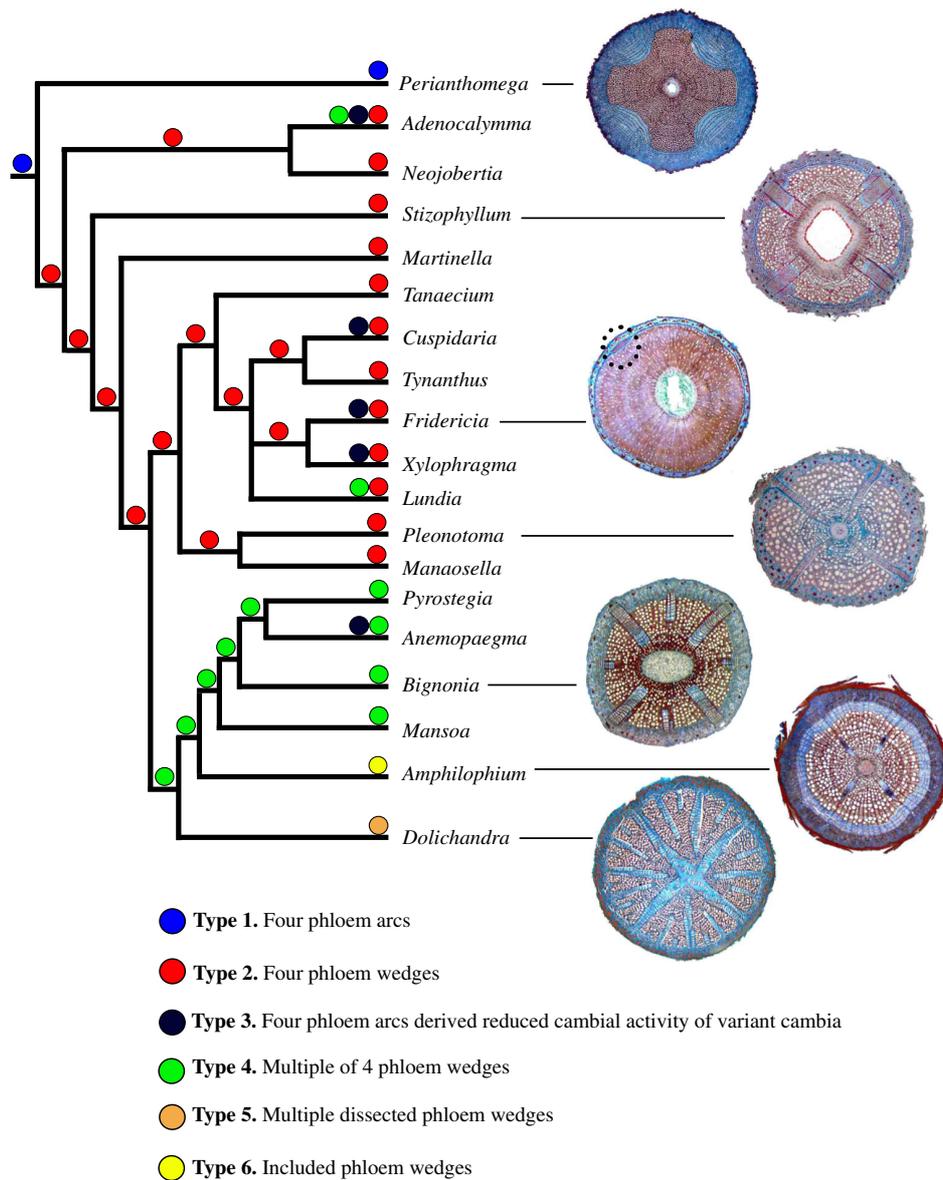
#### Bignoniaceae

We sampled all 21 genera and 49 out of the 393 species currently included in Bignoniaceae (sensu Lohmann and Taylor 2014). Sampling included *Adenocalymma* (8 of 82), *Amphilophium* (5 of 47), *Anemopaegma* (1 of 45), *Bignonia* (4 of 28), *Callichlamys latifolia* (monotypic),

*Cuspidaria* (2 of 19), *Dolichandra* (2 of 8), *Fridericia* (5 of 67), *Lundia* (3 of 13), *Manaosella cordifolia* (monotypic), *Mansoa* (3 of 12), *Martinella* (1 of 2), *Neojobertia* (both species), *Pachyptera* (1 of 4 species), *Perianthomega vellozoi* (monotypic), *Pleonotoma* (3 of 17 species), *Pyrrostegia* (1 of 2 species), *Stizophyllum* (1 of 3 species), *Tanaecium* (3 of 17 species), *Tynanthus* (1 of 15 species), and *Xylophragma* (2 of 7 species).

#### Diagnostic features

Bignoniaceae are characterized by the presence of a cambial variant, denominated furrowed xylem, with four or multiples of four phloem wedges that interrupt the xylem



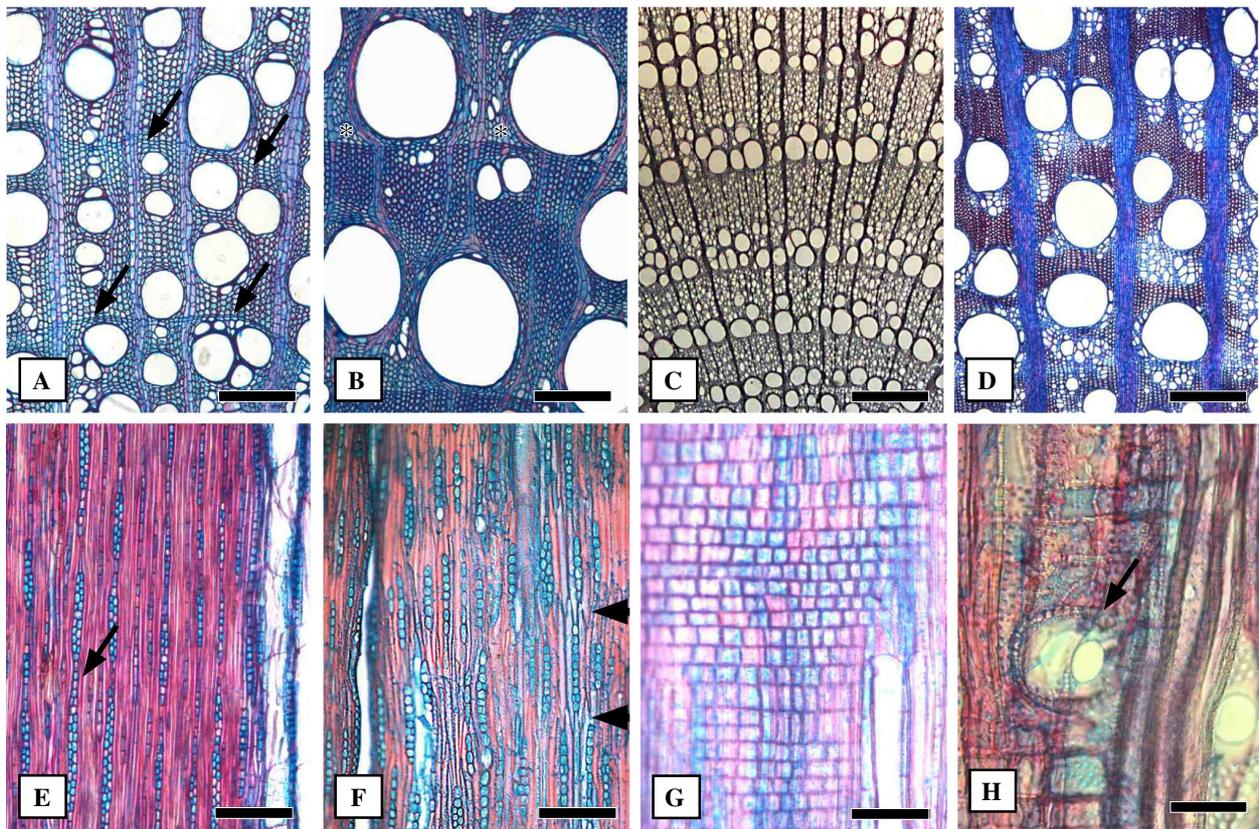
**Fig. 10** Phylogenetic mapping of the six types of cambial variants found in Bignoniaceae; entire stem, as seen in transverse sections. *Type 1* *Perianthomega vellozoi*, four broad phloem arcs. *Type 2* *Stizophyllum riparium* and *Pleonotoma tetraquetra*, four phloem wedges. *Type 3* *Fridericia platyphylla*, four narrow phloem arcs derived from a

reduced activity of the variant cambia, typical of shrub species. *Type 4* *Bignonia binata*, multiple of four phloem wedges. *Type 5* *Dolichandra unguis-cati*, multiple dissected phloem wedges. *Type 6* *Amphilophium crucigerum*, included phloem wedges

(Fig. 10). The wood anatomy of members of Bignoniaceae is characterized by scanty paratracheal to vasicentric axial parenchyma, generally tall (>1 mm) and heterocellular rays, and septate fibers (Fig. 11)

Cambial variants in Bignoniaceae have the conspicuous form of a cross in transverse section (Fig. 10). The stems of Bignoniaceae can be sorted in six types, according to the form and distribution of the phloem arcs/wedges. *Type 1* corresponds to four broad equidistant phloem arcs (Fig. 10) and is exclusive of *Perianthomega*, a monotypic genus that

is sister to all other Bignoniaceae. In this type, the cambium lines the entire stem circumference, without disjunctions. *Type 2* corresponds to four equidistant phloem wedges, formed by the presence of a cambial disjunction and inclusion within the phloem wedges (Fig. 10). *Type 2* is most common in Bignoniaceae, and is present in 12 of the 21 genera of the tribe (Fig. 10). *Type 3* corresponds to four narrow phloem arcs (Fig. 10) derived from delayed development of the phloem wedges and is typical of the shrubby species. This type was recorded in *Adenocalymma*



**Fig. 11** Wood anatomy of Bignoniaceae. **a** *Mansoa difficilis*, TS, diffuse porous wood, growth rings delimited by thick-walled and radially flattened fibers (arrows), vessel dimorphism present. **b** *Tynanthus cognatus*, TS, semi-ring porous wood, delimited by radially flattened fibers and a marginal band of parenchyma, vessel dimorphism present. Wide vessels generally solitary. **c** *Bignonia capreolata*, TS, ring porous wood. **d** *Amphilophium crucigerum*, TS, diffuse porous wood, wide vessels generally solitary, combined with clusters

of narrow vessels, wide rays. **e** *Stizophyllum riparium*, LT, biseriate rays, fusion of rays common (arrow). **f** *Dolichandra unguis-cati*, LT, uniseriate and short rays storied. Fibers also storied (arrows). **g** *Fridericia speciosa*, LR, heterocellular mixed rays. **h** *Neojobertia mirabilis*, LR, perforated ray cell (arrow) with simple perforation plate. Scale bars **a, b, e, f** 200  $\mu\text{m}$ , **c** 800  $\mu\text{m}$ , **d** 300  $\mu\text{m}$ , **g** 150  $\mu\text{m}$ , **h** 40  $\mu\text{m}$

*nodosum*, *A. peregrinum*, *Cuspidaria pulchra*, and *Fridericia platyphylla*, all shrubs growing in the *Cerrado* (Brazilian savannah). The wedges may be very reduced making it impossible to detect without magnification (circled in Fig. 10). *Type 4* corresponds to multiple of four phloem wedges. These species start their development with four phloem wedges, but form additional wedges between the previous ones, always in multiples of four. This type is encountered in the all species from the clade formed by *Amphilophium*, *Anemopaegma*, *Bignonia*, *Dolichandra*, *Mansoa*, and *Pyrostegia*, in most species of *Lundia* and a few *Adenocalymma* (Fig. 10). *Amphilophium* and *Dolichandra*, however, go further in their development generating the next two types to be described. *Type 5* corresponds to the ‘multiple dissected type,’ in which multiple of four phloem wedges are also formed, and in which the proliferation of unligified axial and ray parenchyma dissects the secondary xylem in pieces. The presence of unligified axial and ray

parenchyma in the xylem, which produces this type, is exclusive to *Dolichandra* (Fig. 10). *Type 6* corresponds to the ‘included phloem wedges type,’ in which multiple of four phloem wedges are formed, but are gradually occluded by the regular cambium at the top of the phloem wedges, eventually including the secondary phloem within the secondary xylem. This type is exclusive to *Amphilophium* (Fig. 10).

*Detailed description*

*Growth rings* are distinct and are delimited by a line or band of marginal parenchyma (Fig. 11a, b), very narrow vessels, thicker-walled and radially flattened fibers (Fig. 11a, b) and dilated rays (Fig. 11b). Discontinuous and merging growth rings are common. *Porosity* usually semi-ring porous (Fig. 11b) to diffuse porous (Fig. 11a). The sole species growing in temperate latitudes, *Bignonia*

*capreolata*, has ring-porous wood (Fig. 11c). Vessels exhibit dimorphism, with wide and narrow vessels combined (Fig. 11a-b, d); wide vessels with  $240 \pm 60 \mu\text{m}$ , predominantly solitary in some species (e.g., *Amphilophium crucigerum*, *Stizophyllum riparium*, *Lundia damazii*) to solitary and multiples of 2–3 (Fig. 11a–d),  $2.57 \pm 0.91$  vessels/group. Narrow vessels ( $20 \pm 6 \mu\text{m}$ ) usually have a conspicuous arrangement, such as radial rows, around the wide vessels or in clusters (Fig. 11d). Frequency  $>50/\text{mm}^2$ . Perforation plates are simple. Intervessel pits are medium ( $8\text{--}10 \mu\text{m}$  in diameter). Vessel-ray pits are similar to intervessel pits in size and shape. Fibers are septate in most species, thin- to thick-walled (Fig. 11a, b, d), and with simple pits. Parenchyma is scanty paratracheal in most species (Fig. 11a), except for the clade formed by *Cuspidaria*, *Fridericia*, *Lundia*, *Tanaecium*, *Tynanthus*, and *Xylophragma* (Fig. 11b; the *Arrabidaea* and allies clade in Lohmann 2006) and *Callichlamys latifolia* (not assigned to any clade; Lohmann 2006) in which vasicentric to aliform parenchyma with short confluences and a marginal band of parenchyma are present (Fig. 11b). Rays are heterocellular mixed (Fig. 11g), higher than 1 mm in most species, with variable width, ranging from uniseriate and short (e.g., *Dolichandra unguis-cati*; Fig. 11f), 2–4 cells wide (e.g., *Stizophyllum riparium*; Fig. 11e) and others are wider (Fig. 11d), 5–10 cells wide (e.g., *Amphilophium crucigerum*). The wood under the phloem wedges (variant secondary xylem) has narrower vessels, and rays unicellular and short. An increase in ray width was observed in some species from the stem center towards the bark (e.g. *Amphilophium crucigerum*). Fusion of rays is common in all species (Fig. 11e). Storied structure is absent in most species, but present in the axial, ray parenchyma and fibers of *Dolichandra* (Fig. 11f), and in the fibers of *Perianthomega vellozoi* and all sampled species of *Amphilophium*, and *Mansoa*. Perforated ray cells vary from very abundant (e.g., *Dolichandra unguis-cati*; *Stizophyllum riparium*; Fig. 11h) to rare (e.g., *Perianthomega vellozoi*). Silica is present in the ray cells of *Pachyptera kerere*.

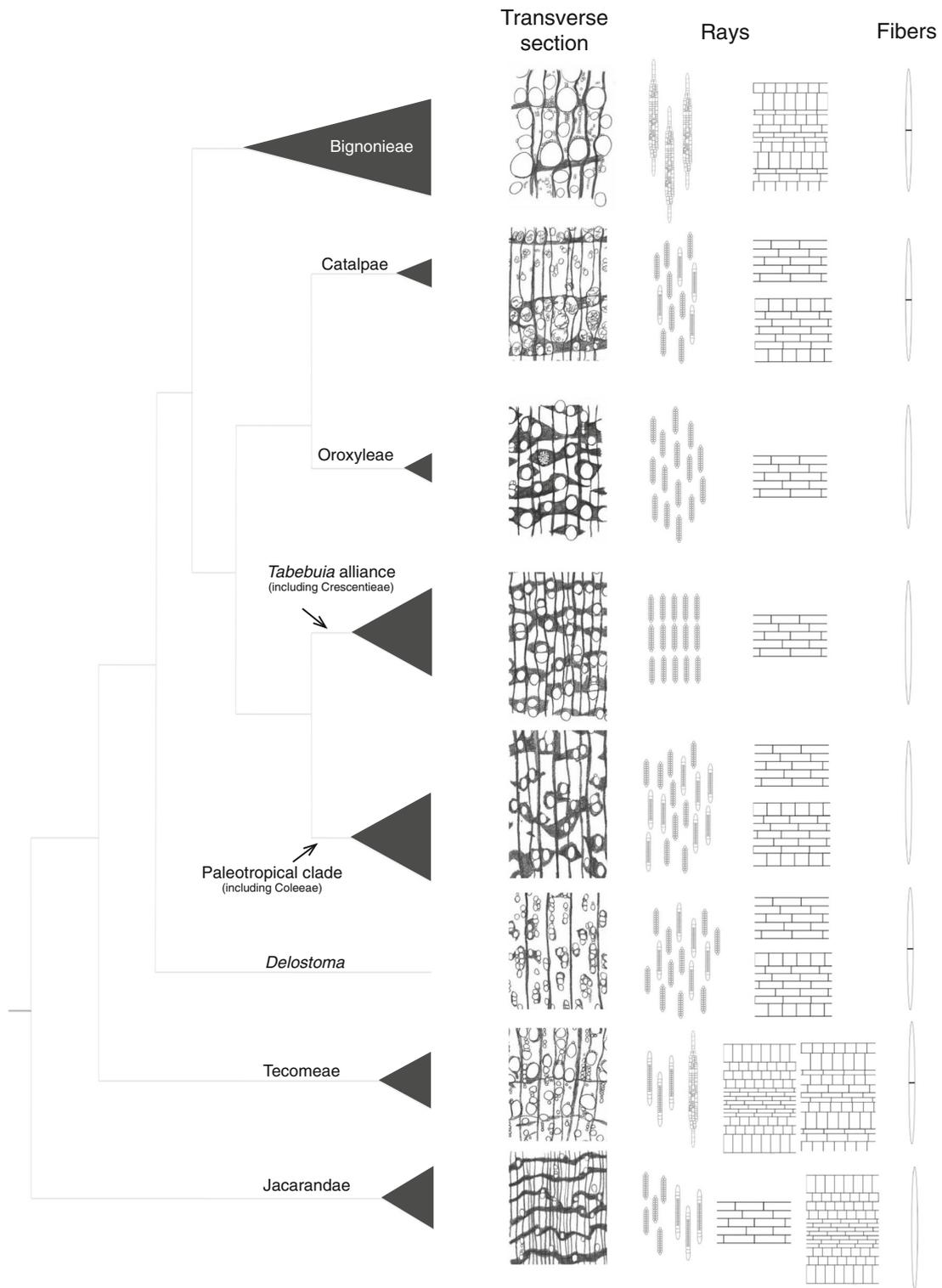
## Discussion

Molecular phylogenetic studies of Bignoniaceae have indicated that several tribes and genera were not monophyletic as traditionally recognized (Spangler and Olmstead 1999; Zjhra et al. 2004; Lohmann 2006; Grose and Olmstead 2007a, b; Li 2008; Olmstead et al. 2009), leading to many changes in the circumscription of taxa within the family. While most of the genus-level clades recognized are well supported by molecular and morphological characters (Grose and Olmstead 2007b; Lohmann and Taylor 2014), several of the higher-level clades (i.e., comparable

to tribal level), however, still lack morphological synapomorphies (Olmstead et al. 2009). In this study, we investigate the wood anatomy of representatives of all major lineages of the Bignoniaceae (except Tourretieae), highlighting the most conspicuous features of each clade (Table 2; Fig. 12), providing a thorough description of their wood anatomy, and suggesting potential synapomorphies (Table 3). The new systematic arrangement of the Bignoniaceae led to more homogeneous and predictable wood anatomical groups (Table 2; Fig. 12). Key features of each clade are summarized and discussed below.

Jacarandaeae were originally described as a tribe by Bentham and Hooker (1876) and later treated as part of Tecomeae s.l. (Gentry 1980; Fischer et al. 2004), but has been resurrected as a tribe and is now known to be sister to all other Bignoniaceae (Olmstead et al. 2009). The two genera of Jacarandaeae, *Digomphia* and *Jacaranda*, share wood anatomical traits that are common to other Bignoniaceae, such as the marginal parenchyma delimiting the growth rings, and paratracheal aliform parenchyma. However, *Digomphia* and *Jacaranda* share the presence of narrow vessels, winged-aliform parenchyma (more conspicuous in *Jacaranda*), and non-storied rays, which supports the circumscription of this tribe. Wood anatomical traits also support the traditional division of *Jacaranda* into sections *Monolobos* and *Dilobos* (Dos Santos and Miller 1997), which were delimited on the basis of number of anther thecae, one or two, respectively. *Jacaranda* section *Monolobos* is characterized by homocellular and uniseriate rays, while *Jacaranda* section *Dilobos* is characterized by heterocellular and multiseriate rays (Dos Santos and Miller 1997). The only exception to this rule is in *Jacaranda copaia*, which is currently assigned to section *Monolobos*, but has anatomical traits that are intermediate between both sections, having multiseriate rays of homocellular composition, and has a unique wood anatomy within the tribe, having the widest vessels in Bignoniaceae trees ( $\sim 300 \mu\text{m}$ ). Additional sampling of *Jacaranda* in molecular phylogenetic studies is still needed in order to test the monophyly of the *Jacaranda* sections as traditionally circumscribed. *Jacaranda* is an ideal genus for further comparative anatomical studies since it presents a great deal of diversity of habits, from underground xylopodial shrubs and subshrubs in the *Cerrado*, treelets, medium-sized to very tall fast-growing trees in forests (e.g. *Jacaranda copaia*), and twigs were shown to represent consistently the wood anatomy of the main trunk wood in the genus (Dos Santos and Miller 1997).

Tecomeae sensu Olmstead et al. (2009) are much more narrowly circumscribed than the tribe Tecomeae previously recognized (Gentry 1992; Fischer et al. 2004). Even though morphological features are still lacking to diagnose this widely distributed tribe (occurring in America, Africa,



**Fig. 12** Drawing summarizing of the most conspicuous wood anatomical features of major Bignoniaceae clades

**Table 3** Potential wood anatomical synapomorphies

Taxa	Anatomical synapomorphy
Jacarandaeae	Axial parenchyma winged-aliform
Catalpeae	Vessel-ray pits simple to semi-bordered
Oroxyleae	Foraminate perforation plates common
Bignoniaceae	Cambial variant: xylem furrowed by four to multiple of four phloem arcs/wedges
<i>Tabebuia</i> alliance	Storied structure
Coleeae	Diffuse axial parenchyma
<i>Amphilophium</i>	Included phloem wedges
<i>Dolichandra</i>	Multiple dissected phloem wedges derived of unlignified parenchyma proliferation
<i>Perianthomega</i>	Four broad phloem arcs

Asia and Oceania; see Olmstead et al. 2009), Tecomeae s.s. are quite homogeneous in wood anatomy (except from *Deplanchea*, whose wood anatomy looks very much like Jacarandaeae), combining features that set aside members of Tecomeae from most other Bignoniaceae. In particular, heterocellular rays with body procumbent and several marginal upright to square cells to heterocellular mixed in some lianas (e.g., *Pandorea jasminoides*), septate fibers, scanty paratracheal parenchyma, and rather narrow vessels (except in the lianas) characterize all members of this clade.

*Delostoma* is a clade of four species of Neotropical treelets and shrubs. Traditionally, *Delostoma* was included within Tecomeae s.l. (Gentry 1980; Fischer et al. 2004), however, *Delostoma*, which bears simple leaves and double calyx, has been difficult to relate to either Paleotropical or Neotropical Bignoniaceae both morphologically (Gentry 1980) and wood anatomically (Dos Santos and Miller 1992). In the most recent phylogeny of Olmstead et al. (2009), *Delostoma* emerged in its own clade, not closely related to any other representatives of Bignoniaceae. Wood anatomically this genus is most similar to Tecomeae s.s., exhibiting heterocellular rays, septate fibers and small vessels usually in radial disposition. It differs from other Tecomeae mainly by the longer tracheary elements (Dos Santos and Miller 1992).

The *Tabebuia* alliance (Olmstead et al. 2009) are composed primarily of Neotropical trees and shrubs with palmately compound leaves; only a few members have simple leaves (e.g., *Crescentia*, *Tabebuia nodosa*). Members of several genera within this clade have invaluable timbers for interior and civil construction (e.g., *Handroanthus*, *Roseodendron*, *Cybistax*, *Paratecoma*). *Tabebuia* s.l., the largest genus within this clade, was shown to be polyphyletic (Grose and Olmstead 2007a) and subdivided in three smaller genera (Grose and Olmstead 2007b):

*Tabebuia* (67 species), *Handroanthus* (30 species), and *Roseodendron* (two species).

Wood anatomical traits strongly support the new generic circumscription. More specifically, the *Tabebuia* Group I proposed by Dos Santos and Miller (1992), which coincides with the Lapacho group of Record and Hess (1943), is characterized by very dense wood and high specific gravity (higher than 0.74), olive brown to blackish heartwood, abundant lapachol in the heartwood vessels, coinciding perfectly with *Handroanthus*. Additional anatomical features of this clade are the large intervessel pits (usually >10 µm), very thick-walled fibers, storied structure and 2–3-seriate rays. The Bignoniaceae species with greatest economical value due to the high quality timber (Record and Hess 1943; Gentry 1992) are now included in this genus. In addition, *Tabebuia* Groups II and III proposed by Dos Santos and Miller (1992) correspond to the newly circumscribed *Tabebuia* s.s. (Grose and Olmstead 2007b). This clade is characterized by the medium basic specific gravity (0.40–0.74), light colored heartwood, not very distinct heartwood and sapwood, and lack of lapachol (Dos Santos and Miller 1992), as well as by small intervessel pits (5–8 µm), 1–2 seriate rays, that are irregularly storied to storied, and thin to thick-walled fibers. Finally, *Tabebuia chrysea* and *Tabebuia donnell-smithii*, two of four species that did not belong to Groups I, II, or III proposed by Dos Santos and Miller (1992), are now treated under *Roseodendron*. The other two species, *Tabebuia nodosa* and *Tabebuia fluviatilis* are also considered quite anomalous anatomically (Dos Santos and Miller 1992) and further phylogenetic data is still needed to confirm their placement among the three segregated genera.

Lapachol (naphthoquinone) was first described in *Tabebuia avellanadae* (= *Handroanthus impetiginosus*) in the XIX century and has antimicrobial properties and numerous applications in pharmacology (Hussain et al. 2007). Inclusion of lapachol in the vessels was evident in dense woods of the *Tabebuia* alliance by Dos Santos and Miller (1992) and us, such as in the genera *Ekmanianthe*, *Godmania*, *Handroanthus* and *Zeyheria*. Studies involving chemical extractions found lapachol in a number of other Bignoniaceae (Hussain et al. 2007), such as *Fernandoa*, *Heterophragma*, *Kigelia*, *Newboldia*, *Phyllarthron*, *Radermachera*, *Stereospermum*, *Tecomella* (Paleotropical clade), *Cybistax*, *Paratecoma peroba* (*Tabebuia* alliance), *Catalpa* (Catalpeae), *Dolichandra unguis-cati* and *Dolichandra quadrivalvis* (Bignoniaceae). Broad chemical extractions of members of the entire family are needed to investigate further the exact distribution of this feature within the Bignoniaceae and its possible taxonomic value.

Tribe Crescentieae, a monophyletic group nested within the *Tabebuia* alliance (Grose and Olmstead 2007a; Olmstead et al. 2009), is also distinct anatomically, especially

due to the abundant aliform confluent parenchyma that forms large bands accounting for almost half of the cells in the wood.

The other seven genera scattered within the *Tabebuia* alliance clade (Olmstead et al. 2009) are much smaller [i.e., *Cybistax* (monotypic), *Ekmanianthe* (ditypic), *Godmania* (ditypic), *Paratecoma* (monotypic), *Sparattosperma* (monotypic), *Spirotecoma* (four species), and *Zeyheria* (ditypic)] and share with the rest of the *Tabebuia* alliance narrow to medium vessels, aliform paratracheal parenchyma, forming short to long confluences, storied or irregularly storied structure (likely an anatomical synapomorphy of the *Tabebuia* alliance; Pace and Angyalossy 2013), exclusively or mostly with homocellular rays and thin to thick-walled non-septate fibers. *Ekmanianthe* was described as having diffuse-in-aggregate axial parenchyma by Gasson and Dobbins (1991); however, neither our specimens nor the ones studied by Dos Santos and Miller (1992) had this feature.

The Paleotropical clade is composed of trees and a few shrubs of pinnately compound leaves (Olmstead et al. 2009), except for members of Coleeae, a tribe endemic to Madagascar, which includes shrubs with simple leaves (Zjhra et al. 2004; Olmstead et al. 2009). Members of this clade are anatomically similar to the *Tabebuia* alliance, except for the absence of storied structure and medium sized vessels.

Members of Coleeae, a monophyletic group nested within the Paleotropical clade are characterized by cauliflory, indehiscent fruits, and pinnately compound leaves (genus *Phylloctenium* with simple leaves and spines; Zjhra et al. 2004). Wood anatomy of Coleeae is unique, with apotracheal diffuse parenchyma and uniseriate rays. Within the tribe, two groups can be recognized by wood anatomy; the first formed by *Colea* and *Ophiocolea*, with thin-walled fibers and abundant apotracheal parenchyma, and the other by *Rhodocolea* and *Phyllarthron*, with very thick-walled fibers and scanty apotracheal parenchyma. The wood anatomical similarities of *Colea* and *Ophiocolea* support the sister group relationship suggested for both genera in the most recent phylogenies of the family (Zjhra et al. 2004; Olmstead et al. 2009).

Oroxyleae were treated initially within Bignoniaceae due to similarities in fruit dehiscence. Oroxyleae were subsequently set apart as a tribe due to the Asian distribution of its members (vs. Neotropical in Bignoniaceae), absence of tendrillate vines, and a stem without cambial variants (Gentry 1980). Oroxyleae have been shown to be a monophyletic group distinct from Bignoniaceae (Spangler and Olmstead 1999; Olmstead et al. 2009), supporting Gentry's proposal. Representatives of Oroxyleae have wood anatomical traits that set them apart, such as the

presence of foraminated perforation plates, homocellular non-storied rays, and lack of cambial variants.

Catalpeae include two genera, *Catalpa* and *Chilopsis*, with *Catalpa* being divided into two sections, *Catalpa* section *Catalpa* and *Catalpa* section *Macrocatappa*. Species of *Catalpa* are disjunctly distributed, with some species occurring in North America (along with *Chilopsis*) and others in east Asia (Li 2008). Catalpeae are rather homogeneous anatomically, except for the absence of semi-ring to ring porous wood and its paratracheal aliform parenchyma with short confluences in the tropical species (e.g., *Catalpa longissima* in section *Macrocatappa*). All species have vessels of medium diameter (130–200 µm), common presence of tyloses, 3-seriate rays, heterocellular, septate fibers in tropical and subtropical species, and the presence of simple to semi-bordered vessel-ray pittings, a novel character typical of this clade (and potential synapomorphy) that has been previously overlooked.

Bignoniaceae are the most species-rich clade of Bignoniaceae, accounting for almost half of the species in the family (Lohmann 2006; Olmstead et al. 2009). Bignoniaceae are clearly monophyletic (Spangler and Olmstead 1999; Lohmann 2006; Olmstead et al. 2009) and unite all Neotropical lianas of the family and a few shrubs (Lohmann 2006). This clade shares a series of unique morpho-anatomical features, such as terminal leaflets modified into tendrils and the presence of a cambial variant (Schenck 1893; Dobbins 1971; Gentry 1980; Dos Santos 1995; Lohmann 2006; Pace et al. 2009; Lohmann and Taylor 2014). The wood of representatives of Bignoniaceae is similar to that of other lianas (Carlquist 1985, 2001) exhibiting vessel dimorphism, heterocellular rays, and cambial variants (Angyalossy et al., in press). The wood of Bignoniaceae differs from lianas of other families by the presence of a generally scanty paratracheal parenchyma, septate fibers, and no vasicentric-tracheids. Parenchyma is only more abundant in the *Arrabidaea* and allies clade (sensu Lohmann 2006), comprising *Cuspidaria*, *Fridericia*, *Lundia*, *Tanaecium*, *Tynanthus* and *Xylophragma*. The only genus not assign to any clade in Lohmann (2006), *Calliclamys latifolia* (monotypic), shares this unusual abundance of parenchyma with members of the *Arrabidaea* and allies clade and might be better placed within this clade, although additional phylogenetic studies are still needed in order to confirm its placement. Storied structure is present in some species, sometimes only in fibers, and sometimes in the axial and ray parenchyma. Silica was found exclusively in one species of Bignoniaceae, *Pachyptera kerere*, and is absent elsewhere in the family. Sampling of the other four species of the genus is needed to determine whether this is a feature exclusive of this species or a potential synapomorphy of the entire genus within the Bignoniaceae.

The shape and distribution of the cambial variants represent synapomorphies of major clades in the tribe (Lohmann 2006) and are of great importance for the wood development of its species in this clade (Lima et al. 2010). Most Bignoniaceae (12 out of 21 genera) have four phloem wedges. Other clades, however, have a different anatomical architecture. *Perianthomega*, for instance, is the only genus with four broad phloem arcs. Three other clades are different by developing multiples of four phloem wedges, namely: *Lundia*, some *Adenocalymma* and a speciose clade that reunites *Amphilophium*, *Anemopaegma*, *Bignonia*, *Dolichandra*, *Mansoa*, and *Pyrostegia*. All these genera develop multiple of four phloem wedges that progress in a predictive manner from four, eight, 16, 32, 64 and so on. *Amphilophium* and *Dolichandra* go through additional developmental changes that are further described below.

*Amphilophium* is characterized by phloem wedges that get included within the secondary xylem (Dos Santos 1995; Pace et al. 2009), an anatomical feature that is derived from the outgrowth of the cambium at the sides of the phloem wedges including these wedges within the xylem (Pace et al. 2009), similarly to what happens in *Strychnos millepunctata* (Loganiaceae; Veenendaal and Den Outer 1993; Angyalossy et al., in press). *Amphilophium* is a newly circumscribed genus (Lohmann and Taylor 2014) that unites all species from six previously recognized genera (Fischer et al. 2004): *Amphilophium*, *Distictella*, *Distictis*, *Glaziovina*, *Haplophium*, and *Pithecoctenium*; most of these were included in the subtribe Pithecocteniinae of Melchior (1927). Mapping the included phloem wedges onto the phylogeny of Bignoniaceae reconstructs this type of cambial variant as ancestral in this tribe (Pace et al. 2009), suggesting it as a potential synapomorphy of the clade and providing further support for the circumscription of this genus.

*Dolichandra* is one of the genera with multiple of four phloem wedges which develop a novel type of cambial variant called “multiple-dissected” phloem wedges (Dos Santos 1995; Pace et al. 2009). In this genus, non-lignified axial and ray parenchyma is present and proliferates during development dissecting the secondary xylem (Pace et al. 2009). *Dolichandra* (sensu Lohmann and Taylor 2014) reunites four genera previously treated as separate: *Dolichandra*, *Macfadyena*, *Melloa* and *Parabignonia* (Fischer et al. 2004). Members of these genera share this unique type of cambial variant (Dos Santos 1995; Lohmann 2006; Pace et al. 2009), supporting the new circumscription of these genera adopted by Lohmann and Taylor (2014). The evolution of cambial variants in Bignoniaceae seems to have involved terminal additions towards complexity enhancement and heterochrony (Pace et al. 2009).

The present study illustrates the diversity of wood anatomy encountered in Bignoniaceae and identifies key

diagnostic character that provide further support to the new circumscription of newly established genera (Grose and Olmstead 2007b; Lohmann and Taylor 2014), tribes and higher-level clades in the family (Olmstead et al. 2009; Table 3). The wood anatomical synapomorphies identified in this study are: (a) Jacarandaeae have a unique winged-aliform parenchyma, (b) Catalpeae are the only tribe where all members have simple to semi-bordered vessel-ray pits, (c) Oroxyleae have foraminated perforation plates common, (d) the *Tabebuia* alliance has storied structure, (e) Coleeae have apotracheal diffuse parenchyma, (f) Bignoniaceae have a cambial variant called furrowed xylem whose types also support genera within the tribe, such as (g) *Amphilophium*, which has included phloem wedges, (h) *Dolichandra*, with multiple-dissected phloem wedges, (i) *Perianthomega* with four broad phloem arcs. These results represent a first step towards a better characterization of Bignoniaceae clades with anatomical characters. These findings highlight the importance of wood anatomical studies as important sources of morphological characters to diagnose major plant clades as a whole.

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## Appendix

Taxa sampled, followed by collector vouchers, wood collections and/or herbaria accession (following index Xylariorum and index Herbariorum) and collection location. The MAD herbarium was incorporated into the WIS herbarium in 2002 but is kept there as a separate collection.

*Adenocalymma bracteatum* DC., *Castanho 153*, Lohmann 861 (SPFw, SPF), Rio Negro, Amazonas, Brazil.

- Adenocalymma comosum** (Cham.) DC., *Pace* 53 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. **Adenocalymma divaricatum** Miers, *Udulutsch* 2808 (SPFw, HRCB), Lençóis, Bahia, Brazil. **Adenocalymma flaviflorum** (Miq.) L.G. Lohmann, *Sousa-Baena* 2 (SPFw, SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. **Adenocalymma neoflavidum** L.G. Lohmann, *Zuntini* 23 (SPFw, SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. **Adenocalymma nodosum** (Silva Manso) L.G. Lohmann, *Pace* 20 (SPFw, SPF), Uberlândia, Minas Gerais, Brazil. **Adenocalymma peregrinum** (Miers) L.G. Lohmann, *Pace* 26 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. **Adenocalymma salmoneum** J.C. Gomes, Lohmann 658 (SPFw, SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. **Amphilophium crucigerum** (L.) L.G. Lohmann, *Pace* 1, *Pace* 2, *Pace* 3, *Pace* 34 (SPFw, SPF), São Paulo, São Paulo, Brazil. **Amphilophium elongatum** (Vahl) L.G. Lohmann, *Pace* 45 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. **Amphilophium magnoliifolium** (Kunth) L.G. Lohmann, *Lohmann* 851 (SPFw, SPF), Rio Negro, Amazonas, Brazil; *Dos Santos* 272 (MADw, MAD, MO, MG), Porto de Moz, Pará, Brazil. **Amphilophium paniculatum** (L.) Kunth, *Pace* 46 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. **Amphilophium pulverulentum** (Sandwith) L.G. Lohmann, *Dos Santos* 279 (MADw, MAD, MO, MG), Senador Jose Porfirio (Sozel), Pará, Brazil. **Amphitecna latifolia** (Mill.) A.H. Gentry, *Fairchild Tropical Garden* x-3-369, Florida, USA. **Amphitecna regalis** (Linden) A.H. Gentry, *Nee & Taylor* 29900, Las Choapas, 5 km Nw of El Doce, Uxpanapa Region, Veracruz, Mexico. **Anemopaegma chamberlaynii** (Sims) Bureau & K. Schum., *Zuntini* 15 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. **Bignonia campanulata** Cham., *Pace* 39 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. **Bignonia capreolata** L., *Nogle* s.n. (MADw), Norfolk County, Dismal swamp, Virginia, USA; *Wilson* 19 (MADw, F), Cow Creek, Texas, USA. **Bignonia magnifica** W. Bull, *Pace* 51 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. **Bignonia priurei** DC., *Zuntini* 13 (SPFw, SPF), Linhares, Espírito Santo, Brazil; *Dos Santos* 87 (MADw, MAD, MO, MG), Marabá, Pará, Brazil. **Callichlamys latifolia** (Rich.) K. Schum, *Zuntini* 175 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil.; *Pace* 42 (SPFw), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil; *Pace* 63 (SPFw, SPF), Ducke Forest Reserve, Manaus, Amazonas, Brazil; **Campsis radicans** (L.) Seem., *Hicoek* 841 (SJRw, Y), Connecticut, USA; *Pond* 448 (MADw, MAD), Camden County, Dismal Swamp, North Carolina, USA. **Catalpa bignonioides** Walter, *Erdman & DeVall* s.n. (MADw, MAD, NY), Gainesville, Florida, USA. **Catalpa longissima** (Jacq.) Dum.Cours., *Pimentel & Garcia* 965 (SJRw, NCI), San Cristóbal, El Tablazo, Nigua riverside, Cordillera central, Dominican Republic; *Collector unknown* s.n. (USw2942, US), Hispaniola Island. **Catalpa speciosa** (Warder ex Barney) Warder ex Engelm., *Collector unknown* s.n. (SJRw, RBHw3217), location unknown; *Collector unknown* s.n. (SJRw, Uw17977), location unknown. **Catophractes alexandri** D. Don, *Dechamps* 1219 (MADw, Tw, MAD), Mocamedes, Angola. **Chilopsis linearis** (Cav.) Sweet, *Pidgeon* s.n. (SJRw, WIS), Otero County, New Mexico, USA; *Johnson* s.n. (MADw, BWCw), Campaign Wash, Arizona, USA; *Collector unknown* s.n. (Kw), location unknown. **Colea gentryi** M. L. Zhjra, *Zhjra* 714 (MADw, WIS), Masoala Peninsula, Madagascar, Africa. **Colea resupinata** M. L. Zhjra, *Zhjra* 785 (MADw, GAS holotype, TAN isotype, WIS), Vokoanina watershed, Masoala Peninsula, Antsiranana, Madagascar. **Crescentia alata** Kunth, *Wiemann & Lemckert* 23 (MADw, CR, LSU), Cañas, Guanacate, Costa Rica; *Collector unknown* s.n. (SJRw), Cuastecomate, Mexico; *Ortega* 12 (USw), Sinaloa, Mexico. **Crescentia kujete** L., *Pace* 80, São Paulo, São Paulo, Brazil; *Dugand* 149 (MADw, SJRW, MAD), Totuma, Colombia. **Cuspidaria pulchra** (Cham.) L.G. Lohmann, *Pace* 24 (SPFw, SPF), Uberlândia, Minas Gerais, Brazil. **Cydistax antisiphilitica** (Mart.) Mart., *Reitz & Klein* 7354 (MADw, HBRw, HBR), Salto do Pilão, Lontras, Santa Catarina, Brazil; *Collector unknown* s.n. (BWCw, SJRW42602), São Paulo, Brazil. **Delostoma integrifolium** D. Don, *Acosta-Solis* 6694 (MADw, SJRW, MAD, F), Limón, Bolívar, Ecuador; *Acosta-Solis* 11648-A (MADw, F), Ecuador. **Deplanchea bancana** (Scheff.) Steenis, *Lai et al.* 68559 (Kw, K), Sarawak, Malaysia; *Forest Department of Java* 2751 (SJRw, L), Menjabung, Dutch East Indies. **Digomphia densicoma** (Mart. ex DC.) Pilg., *Nee* 31168 (MADw, VEN, NY), Cerro de la neblina, Amazonas, Venezuela; *Maguire* 28311 (BWCw, USw), Mérida, Venezuela. **Dolichandra unguiculata** (Vell.) L.G. Lohmann, *Zuntini* 176 (SPFw, SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. **Dolichandra unguis-cati** (L.) L.G. Lohmann, *Ceccantini* 2687 (SPFw, SPF), Matozinhos, Minas Gerais, Brazil; *Groppo* 322 (SPF), São Paulo, São Paulo, Brazil. **Dolichandra quadrivalvis** (Jacq.) L.G. Lohmann, *Gentry* 58691 (MADw, MO), São Paulo, São Paulo, Brazil. **Dolichandrone atrovirens** (Roth) K. Schum., *Brown* s.n. (DDw, DD), Dehradun, India; *Collector unknown* s.n. (SJRw), Myanmar.; *Kanehira* 132 (SJRw), Palau, Micronesia. **Ekmanianthe actinophylla** (Griseb.) Urb., *Fors* 11 (MADw, SJRW, MAD), Havana, Cuba; *Leon* 14358 (SJRw, NY), Cuba. **Fernandoa adenophylla** (Wall. ex G. Don) Steenis, *collector unknown* s.n. (Kw 108, 427, 433, 435), location unknown. **Fernandoa**

- magnifica* Seem, *Schlieben* 459 (SJRw, MAD), Tanganyika, Tanzania. *Fridericia chica* (Bonpl.) L.G. Lohmann, *Pace* 50 (SPFw), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. *Fridericia conjugata* (Vell.) L.G. Lohmann, *Pace* 44 (SPFw), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. *Fridericia platyphylla* (Cham.) L.G. Lohmann, *Pace* 22, *Pace* 23 (SPFw, SPF), Uberlândia, Minas Gerais, Brazil. *Fridericia samydoides* (Cham.) L.G. Lohmann, *Pace* 49 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. *Fridericia speciosa* Mart., *Pace* 40 (SPFw, SPF), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. *Godmania aesculifolia* (Kunth) Standl., *Breedlove* 9563 (MADw, DS), Chiapas, Mexico; *Williams* 10233 (MADw, F), Aragua, Venezuela; *Smith* 3368 (SJRw, MAD), British Guiana. *Handroanthus barbatus* (E.Mey.) Mattos, Loureiro s.n. (BCTw11727, INPA); *Maguire* 41572 (SJRw, NY), Rio Pacimoni-Yatua, Venezuela. *Handroanthus chrysotrichus* (Mart. ex DC.) Mattos, *Pinho* 6 (BCTw, SP), São Simão, São Paulo, Brazil; *Pace* 188, 190 (SPFw, SPF), São Paulo, São Paulo, Brazil. *Handroanthus impetiginosus* (Mart. ex DC.) Mattos, *Ducke* 363 (SJRw, MAD), Brazil; *Pinho* 2 (BCTw, SP), São Simão, São Paulo, Brazil. *Handroanthus serratifolius* (Vahl) S.O.Grose, *Lima* s.n. (BCTw), Pará, Brazil; *Silva* 3281 (BCTw, INPA), Jari, Pará, Brazil. *Heterophragma quadriloculare* (Roxb.) K.Schum., *Brown* s.n. (SJRw, DDw1106), India; *Dehra Dun* s.n. (SJRw, DDw 241), India; *Pearson* s.n. (MADw), India. *Heterophragma sulfureum* Kurz, *Conservator of Forests* 1238 (SJRw), Burma. *Jacaranda brasiliiana* Lam., *Collector unknown* s.n. (FPBw1755), Brazil; *Collector unknown* s.n. (SJRw, MAD), Brazil. *Jacaranda copaia* (Aubl.) D.Don, *Cabrera* 41, 42 (MADw, MAD), Puerto Carare, Santander, Colombia. *Jacaranda obtusifolia* Bonpl., *Conservator of Forests* 2049 (SJRw), British Guiana; *Smith* 3125 (SJRw, MAD), British Guiana. *Jacaranda puberula* Cham., *Hoehne* 28168 (SJRw, MAD), Brazil; *Reitz* 14198 (BWCw, MAD), Santa Catarina, Brazil. *Jacaranda ulei* Bureau & K.Schum., *Dos Santos* 167, 168 (MADw, MAD, MO, MG), Parauapebas, Pará, Brazil. *Kigelia africana* (Lam.) Benth., *Schlieben* 368 (SJRw, MAD), Tanganyika, Democratic Republic of Congo. *Lundia damazii* C. DC., *Pace* 55, *Pace* 56 (SPFw, SPF), São Paulo, São Paulo, Brazil. *Lundia glazioviana* Kraenzl., *Zuntini* 126 (SPFw, SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. *Lundia longa* (Vell.) DC., *Zuntini* 1 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil; *Pace* 227 (SPFw, SPF), Biological Reserve of Poço das Antas, Rio de Janeiro, Brazil. *Manaosella cordifolia* (DC.) A.H. Gentry, *Pace* 41 (SPFw, SPF), Brazil, Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil; *Dos Santos* 88 (MADw, MAD, MO, MG), Marabá, Rio Doce S.A. Forest reserve, 48 km from Marabá, Pará, Brazil; *Dos Santos* 308 (MADw, MAD, MO, MG), Senador José Porfírio, Xingu riverside. *Mansoa difficilis* (Cham.) Bureau & K. Schum., *Pace* 35 (SPFw), São Paulo, São Paulo, Brazil; *Zuntini* 4 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. *Mansoa onohualcoides* A.H. Gentry, *Zuntini* 276 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. *Mansoa standleyi* (Steerm.) A.H. Gentry, *Pace* 43 (SPFw), Living collection Plantarum Institute, Nova Odessa, São Paulo, Brazil. *Markhamia lutea* (Benth.) K.Schum., *collector unknown* s.n. (Kw525), Equatorial Guinea. *Markhamia stipulata* (Wall.) Seem., *collector unknown* s.n. (Kw440), Thailand. *Martinella obovata* (Kunth) Bureau & K. Schum., *Zuntini* 7 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil; *Dos Santos* 237 (MADw, MAD, MO, MG), Porto de Moz, Xingu riverside, Pará, Brazil; *Dos Santos* 317 (MADw, MAD, MO, MG), Gurupa, Moju riverside, tributary of the Amazon river, Pará, Brazil. *Millingtonia hortensis* L.f., *van Beusekom* 3426 (TWTw, L), Saeat Kanchanaburi, Thailand; *Brown* 3160 (SJRw, DDw), India; *collector unknown* s.n. (Kw), Thailand. *Neojobertia mirabilis* (Sandwith) L.G. Lohmann, *Dos Santos* 48 (MADw, MAD, MO, MG), Buriticupu Forest Reserve, Maranhão, Brazil. *Neojobertia* sp. nov., *Zuntini* 18, Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. *Newbouldia laevis* (P.Beauv.) Seem., *Vigne* 1722 (SJRw, MAD), Gold Coast, Nsuta, Ghana. *Ophiocolea floribunda* (Bojer ex Lindl.) H.Perrier *Zhira* s.n. (MADw, MAD), Masoala Peninsula, Madagascar. *Oroxylum indicum* (L.) Kurtz, *China Academy of Forestry* s.n. (TWTw7424, CAFw13841), China; *Jacobs* 8493 (TWTw, L), Lampung, Sumatra; *Kanehira* s.n. (TWTw, FUw B.401), Java, Indonesia; *Brown* 1179 (SJRw, DDw), India. *Pachyptera kerere* (Aubl.) Sandwith, *Castanho* 143, *Lohmann* 834 (SPF), Negro riverside, Amazonas, Brazil; *Santos* 226 (MADw, MAD, MO, MG), Melgaco, Marajó Island; Mappari riverside, Pará, Brazil; *Dos Santos* 274 (MADw, MAD, MO, MG), Porto de Moz; Xingu riverside near the Açaí river, Pará, Brazil; *Dos Santos* 291 (MADw, MAD, MO, MG), Senador José Porfírio, Xingu river near foz do Igarapé Guará; *Dos Santos* 292 (MADw, MAD, MO, MG), Marabá, Rio Doce S.A. Forest reserve, Sororó riverside. *Pajanelia longifolia* (Willd.) K.Schum., *Conservator of Forests* 8188 (SJRw), Rangoon, Burma; *collector unknown* s.n. (Kw528), Malaya, Malaysia. *Pandorea jasminoides* (Lindl.) K.Schum., *Pace* 18, 19 (SPFw, SPF), Cultivated in Campinas, São Paulo, Brazil. *Paratecoma peroba* (Record) Kuhl., *Castro* 284, 578 (BCTw), Rio Doce, Espírito Santo, Brazil. *Parmentiera cereifera* Seem., *Curtis* s.n. Fairchild Bot. Gard. X4- 183 (MADw), Florida, USA. *Parmentiera macrophylla* Standl., *Cooper* 402 (MADw, SJRW, MAD), Panama; *Stork* 1894 (SJRw,

MAD), Costa Rica. *Perianthomega velozoi* Bureau, Pace 10, Pace 15 (SPFw, SPF), Mata do Paraíso, Viçosa, Minas Gerais, Brazil; Pace 28, Pace 29 (SPFw, SPF), Santa Cruz de la Sierra, Santa Cruz, Bolivia. *Phyllarthron bojeranum* DC., G 31 (SJRw, CTFw), Region Cotier Est, Madagascar. *Pleonotoma melioides* (S. Moore) A.H. Gentry, Dos Santos 174 (MADw, MAD, MO, MG), Parauapebas, Serra dos Carajas Biological Reserve; Dos Santos 298 (MADw, MAD, MO, MG), Senador José Pontifício, Pará, Brazil. *Pleonotoma stichadenia* K. Schum., Zuntini 7 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil; Dos Santos 187 (MADw, MAD, MO, MG), Parauapebas, Pará, Brazil. *Pleonotoma tetraquetra* (Cham.) Bureau, Ozório-Filho 11, São Paulo, São Paulo, Brazil. *Podranea ricasoliana* (Tanfani) Sprague, Pace 11 (SPF), São Paulo, São Paulo, Brazil. *Pyrostegia venusta* (Ker Gawl.) Miers, Pace 17 (SPFw, SPF), Campinas, São Paulo, Brazil; Pace 36 (SPFw, SPF), São Paulo, São Paulo, Brazil. *Radermachera ignea* (Kurz) Steenis, Conservation of Forests 2444 (SJRw), Burma. *Radermachera gigantea* (Blume) Miq., Van de Koppel 4780 (SJRw, L), Java, Indonesia. *Radermachera glandulosa* (Blume) Miq., Janssonius 1214 g (SJRw), Java, Indonesia; collector unknown s.n. (Kw), Burma. *Radermachera pinnata* (Blanco) Seem., Philippine Bureau of Forestry 342 (SJRw), Phillipines. *Radermachera sinica* (Hance) Hemsl., NTU 408, Taiwan. *Rhodocolea multiflora* Zhjra, Zhjra 836, Vokonina, Masoala Peninsula, Madagascar. *Rhodocolea nycteriphilla* Zhjra, Zhjra 810, Vokonina, Masoala Peninsula, Madagascar. *Rhodocolea telfairae* (Bojer ex Hook.) H.Perrier, collector unknown s.n. (SJRw10766), Madagascar. *Roseodendron donnell-smithii* (Rose) Miranda, Williams 8734 (MADw, F), Fortuno, Coatzacoalcos River, Veracruz, Mexico; Williams 9382 (MADw, F), Ubero, Oaxaca, Mexico; William 9458 (MADw, F), Mexico; collector unknown s.n. (Kw920), Venezuela. *Sparattosperma leucanthum* (Vell.) K.Schum., collector unknown s.n. (BCTw 2486), Brasília, Distrito Federal, Brazil. *Spathodea campanulata* P.Beauv., Chevalier 140 (SJRw, K), Gabon; collector unknown s.n. (Kw529), Uganda. *Spirotecoma spiralis* (C.Wright ex Griseb.) Pichon, Bucher 90 (SJRw, MAD), Cuba. *Stereospermum chelonoides* (L.f.) DC., Istituto Botanico dell'Università di Firenze 706 (BCTw), India. *Stizophyllum riparium* (Kunth) Sandwith, Pace 16, Pace 33 (SPFw, SPF), São Paulo, São Paulo, Brazil; Zuntini 9 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. *Tabebuia aurea* (Silva Manso) Benth. & Hook.f. ex S.Moore, Gerolamo 3 (SPFw), São Paulo, São Paulo, Brazil. *Tabebuia cassinoides* (Lam.) DC., Williams 13809 (MADw, F), Puerto Ayacucho, Amazonas, Venezuela. *Tabebuia fluviatilis* (Aubl.) DC., Lobato 447 (MADw, MGw, MG), Barcarena, Pará, Brazil; Conservation of Forests 4071 (SJRw), British Guiana; collector

unknown (SJRw12019), location unknown). *Tabebuia heterophylla* (DC.) Britton, Dugand 33754 (BCTw), Colombia. *Tabebuia obtusifolia* (Cham.) Bureau, Kuhlmann (BCTw, RB), Espírito Santo, Brazil. *Tabebuia rigida* Urb., Instituto de Tecnologia do Rio Grande do Sul s.n. (BCTw), Rio Grande do Sul, Brazil. *Tabebuia rosealba* (Ridl.) Sandwith, CVRD Morais Jesus s.n. (BCTw), Linhares, Espírito Santo, Brazil. *Tanaecium bilabiatum* (Sprague) L.G. Lohmann, Lohmann 850 (SPF), Rio Negro Amazonas, Brazil. *Tanaecium duckei* (A.Samp.) A.H.Gentry, Dos Santos 179, 186, Serra do Carajás Biological Reserve, Companhia Vale do Rio Doce, Parauapebas, Pará, Brazil. *Tanaecium pyramidatum* (Rich.) L.G. Lohmann, Pace 14, Pace 35 (SPFw, SPF), São Paulo, São Paulo, Brazil; Dos Santos 101 (MADw, MAD, MO, MG), Marabá, Rio Doce S.A. forest reserve, Sororó riverside, Pará, Brazil. *Tecoma cochabambensis* (Herzog) Sandwith, Salomon 6684 (MADw, MO) Murillo, La Paz, Bolivia. *Tecoma fulva* (Cav.) G.Don, collector unknown s.n. (SJRw32082), location unknown. *Tecoma stans* (L.) Juss. ex Kunth, Jack 5693 (SJRw, MAD), Santa Clara, Belmonte, Cuba; Dugand 216 (SJRw, MAD), Colombia; Williams 12254 (MADw, F), Federal District, Venezuela. Pace 422, 423 (SPFw, SPF, MO), Magdalena Ocotlán, Oaxaca, Mexico. *Tecomaria capensis* (Thunb.) Spach, Rimbach 832 (SJRw, MAD), Ecuador. *Tecomella undulata* (Sm.) Seem., collector unknown s.n. (Kw550), location unknown. *Tynanthus cognatus* (Cham.) Miers, Pace 9a, Pace 9b (SPFw, SPF), São Paulo, São Paulo, Brazil. *Xylophragma myrianthum* (Cham. ex Steud.) Sprague, Zuntini 263 (SPF), Vale do Rio Doce Forest Reserve, Espírito Santo, Brazil. *Xylophragma pratense* (Bureau & K.Schum.) Sprague, Dos Santos 140 (MADw, MAD, MO, MG), Marabá, Rio Doce S.A. forest reserve, Pará, Brazil. *Zeyheria montana* Mart., Pacheco 2762 (SJRw), Minas Gerais, Brazil; Heringen 4130 (BCTw, MADw), Rio de Janeiro, Brazil. *Zeyheria tuberculosa* (Vell.) Bureau ex Verl., Schmidt 143 (SJRw, M), Bolivia.

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